

**Centro Congressi Frentani
Workshop
“Il fotovoltaico in Italia nel triennio 2011-2013”**

Relatori : D.ssa Roberta Congestri – Dott. Ing. Marcello Diano

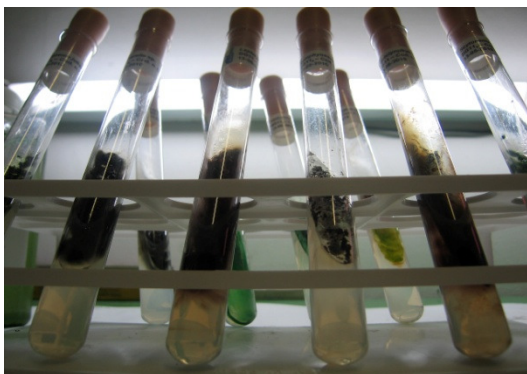
Contenuti: Le biomasse algali: una potenziale opportunità
per l'evoluzione delle serre fotovoltaiche



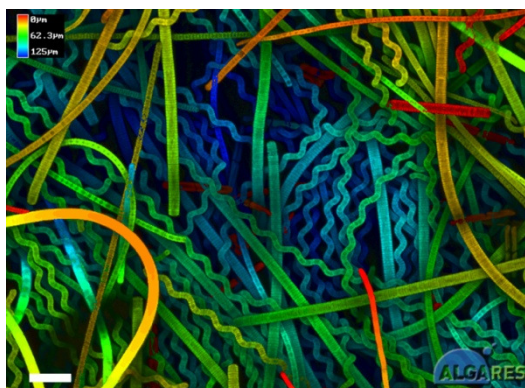
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“Il fotovoltaico in Italia nel triennio 2011-2013”**

Relatori : D.ssa Roberta Congestri - Università di Roma “Tor Vergata” e AlgaRes

Contenuti: Biomasse algali ed energia



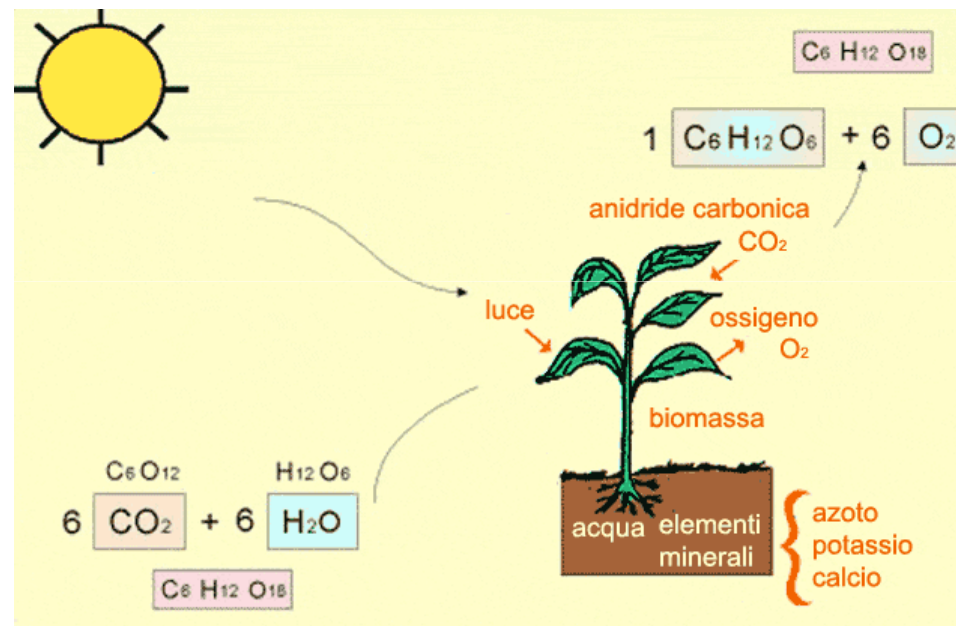
Biomasse
Biomolecole
Trattamento reflui



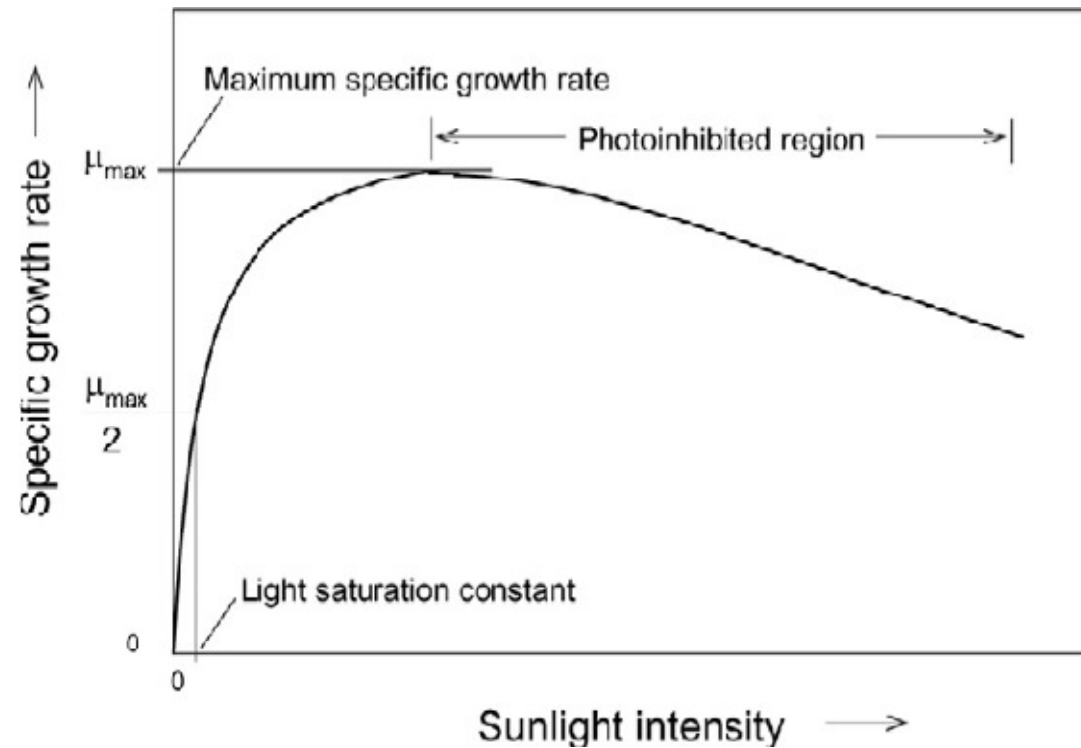
Controllo qualità ambientale
Conservazione beni culturali

Alghe ed energia?

Sfruttare l'energia del sole attraverso la fotosintesi delle alghe

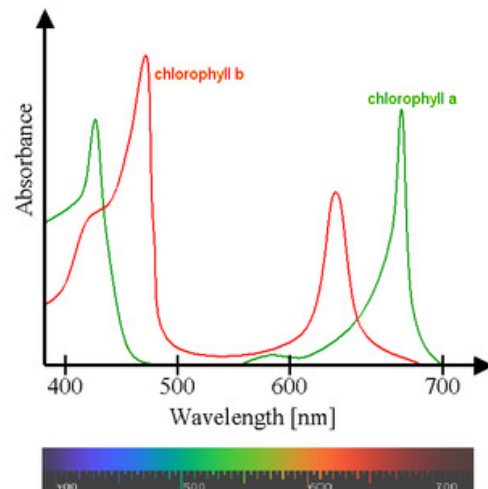
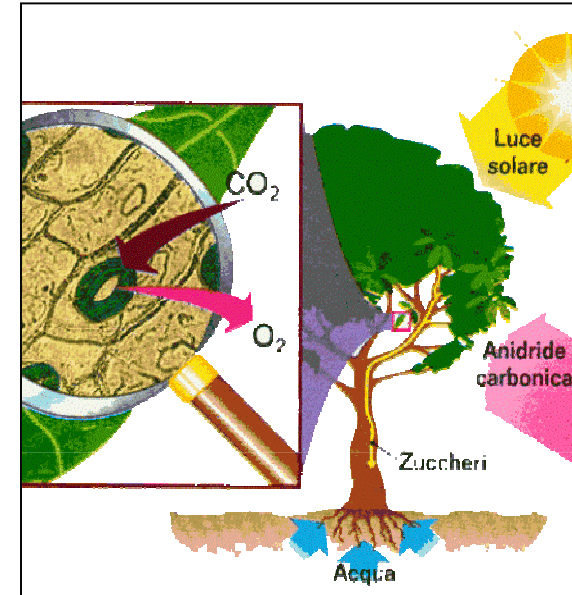
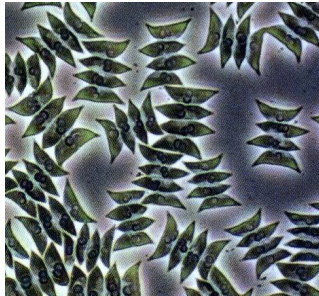
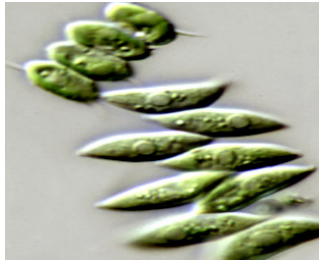


- processo “inventato” **una** sola volta nel corso dell’evoluzione dei vegetali, dalle **alghe!!!**
- > 3 miliardi di anni fa
- conversione E solare in E biochimica (biomassa, molecole ad alta energia...)
- ottimizzazione “naturale”

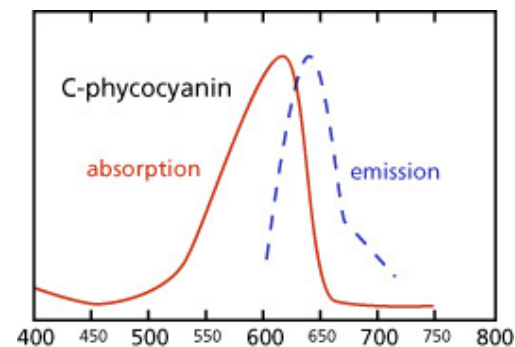


Effect of light intensity on specific growth rate of microalgae, Chisti 2007

Efficienza conversione energia solare: alghe vs piante



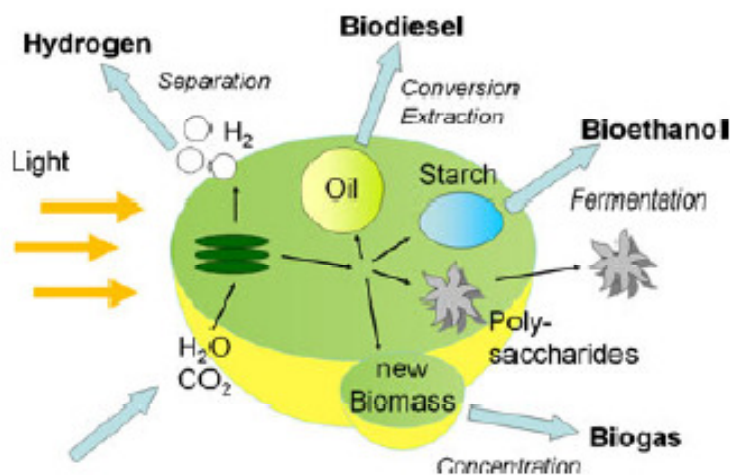
Pigmenti



CO₂

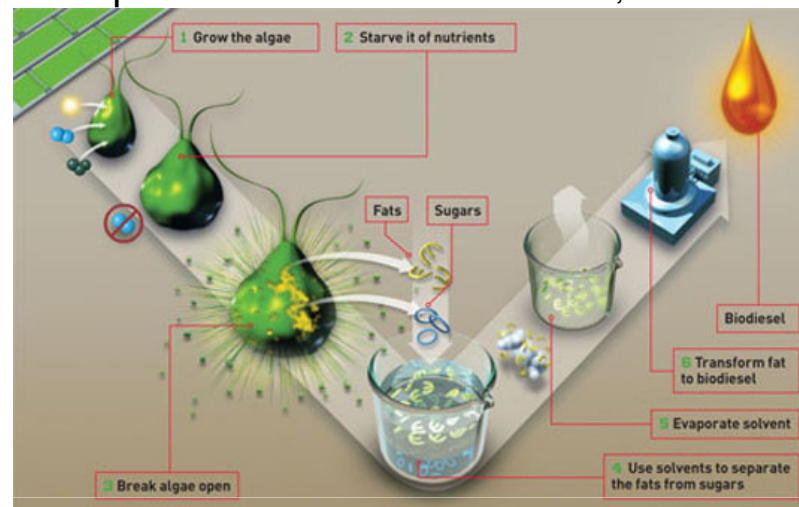


Generazione di energia dalle alghe

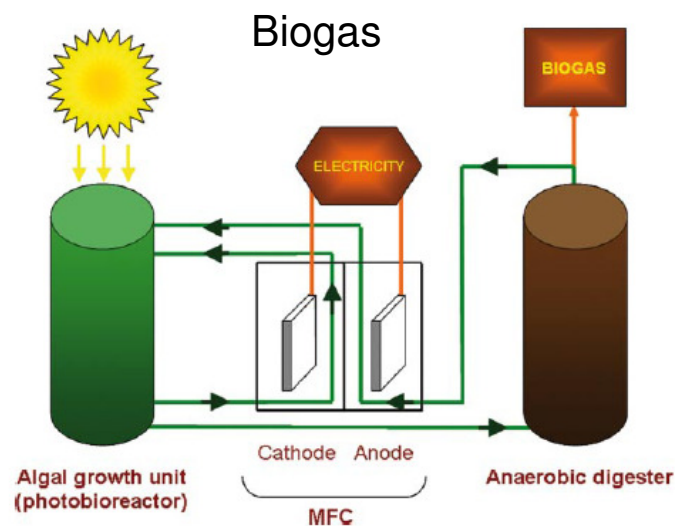


Possible routes to energy products

Oli per biocarburanti: biodiesel , kerosene..



Pigmenti per celle solari organiche



Perché le alghe?

Nessuna competizione con risorse alimentari

Elevato tasso di crescita

Elevato contenuto in lipidi

Inferiore richiesta acqua

Manipolazione della produzione di acidi grassi (condizioni colturali)

Non richiedono uso di pesticidi o erbicidi

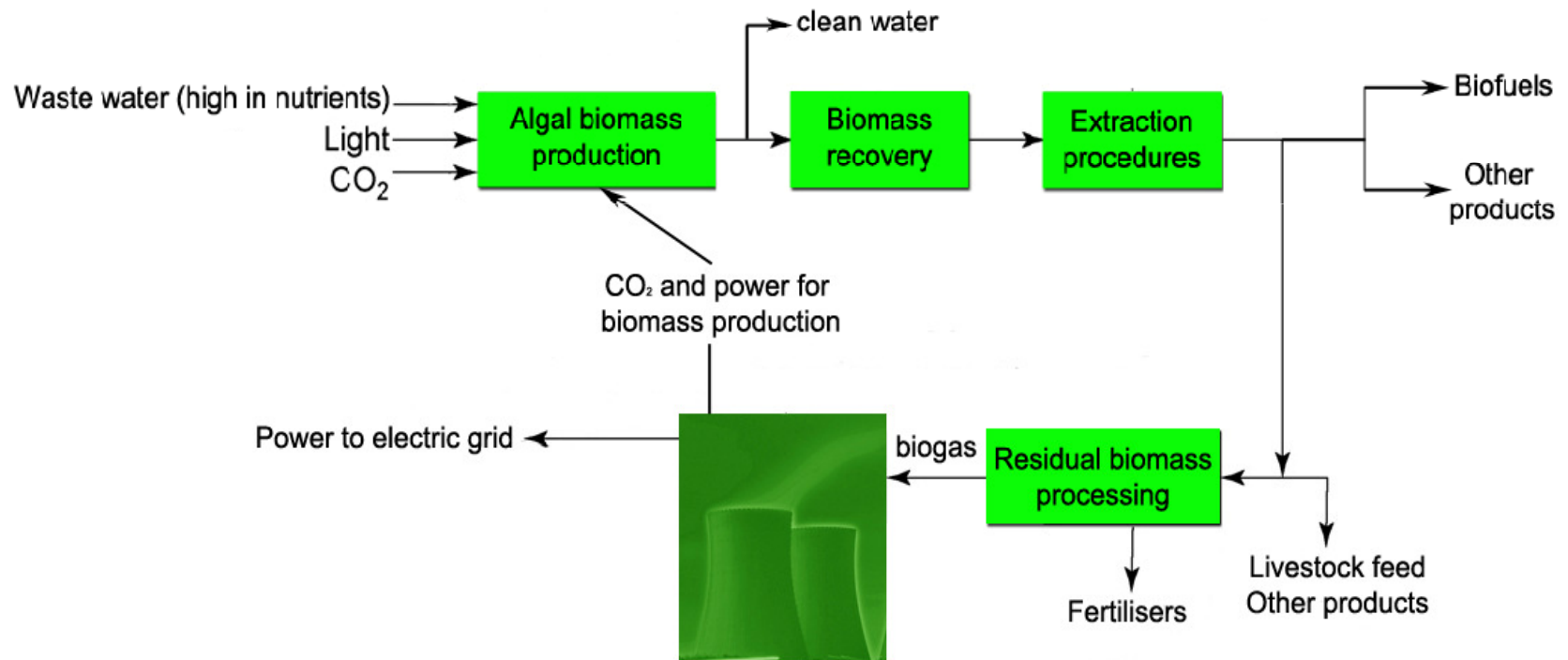
Riutilizzo di terreni non coltivabili o contaminati

Utilizzo acque reflue, ricche in azoto e fosforo, per colture intensive

Utilizzo della biomassa per estrazione composti

Incentivazione altre applicazioni commerciali

PROCESSO AUTOSOSTENIBILE DI PRODUZIONE DI BIOMASSA ALGALE A BASSO IMPATTO AMBIENTALE



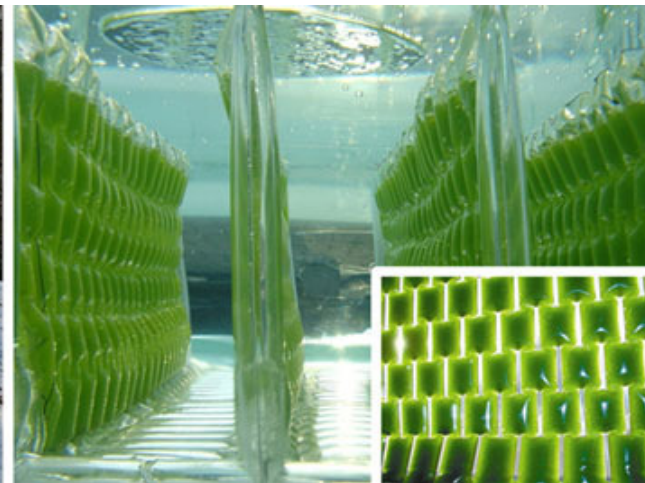
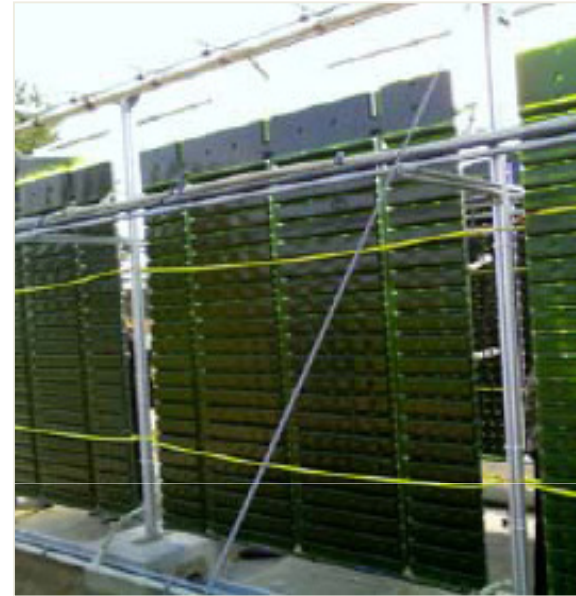
A scala industriale: due sistemi di coltura per la crescita delle microalghe aperti



OPEN PONDS



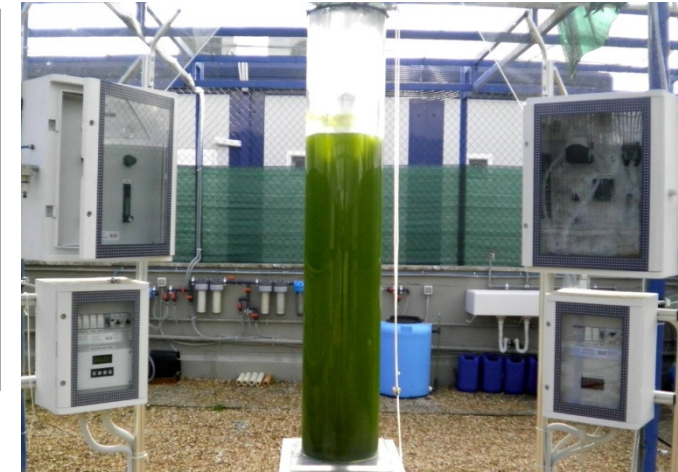
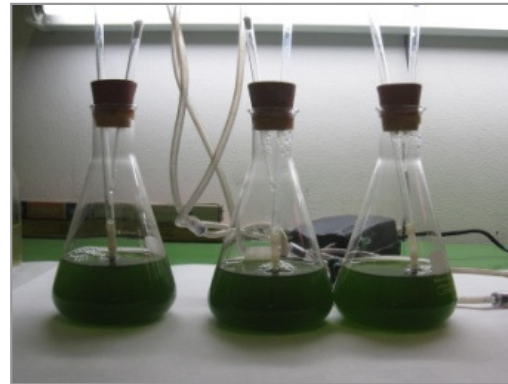
chiusi



FOTOBIOREATTORI

Morweiser et al. 2010

In questo scenario... screening ceppi in lab e in prototipo PBR



T = 18 °C
PPFD = 18 $\mu\text{mol photon m}^{-2}\text{s}^{-1}$
L/D cycle = 14:10

***N. oculata* CCAP849/1**
Coccal green FIU-1
Filamentous cyanos FIU-1, 2
CA2a-1

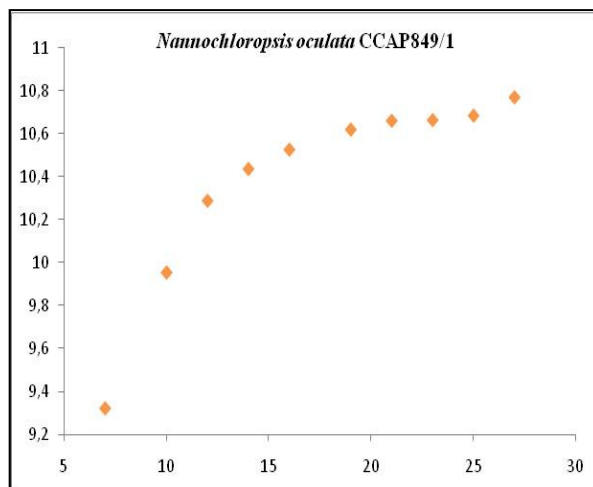
***N. oculata* CCAP849/1**
Coccal green FIU-1

T = ambient, 25 °C
Solar irradiance
L/D cycle seasonal

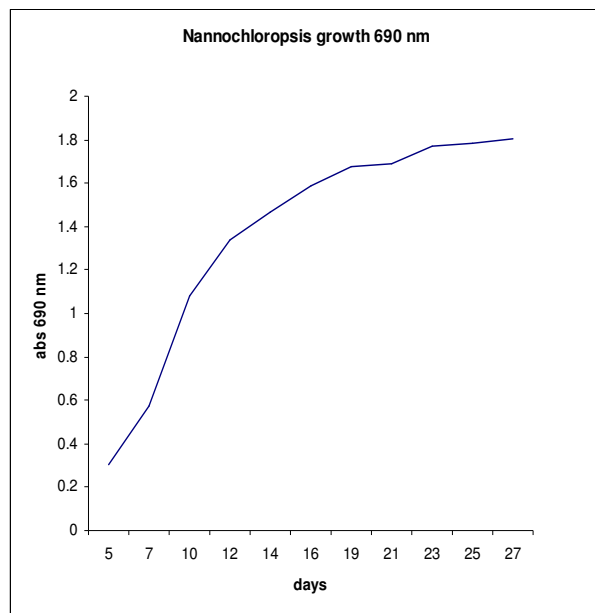
***N. oculata* CCAP849/1**
Coccal green FIU-1
Filamentous cyanos FIU-1, 2
Raphid diatom

Lab

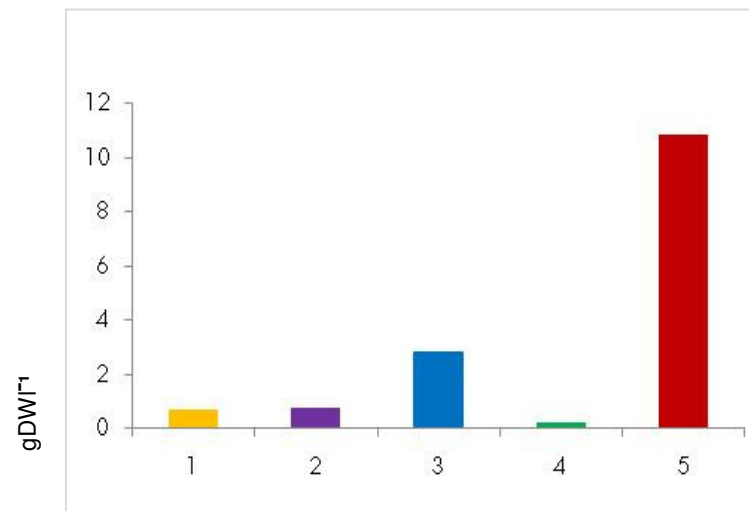
Curve di crescita



PBR



Produttività



AlgaSeed: Inoculi

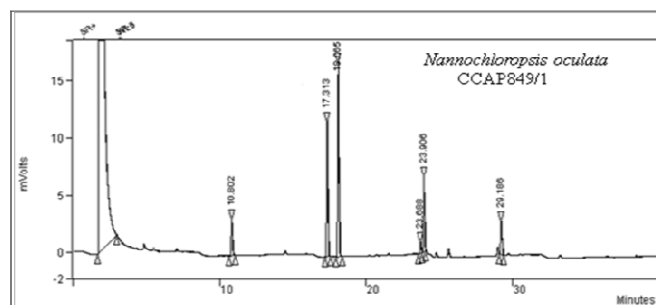
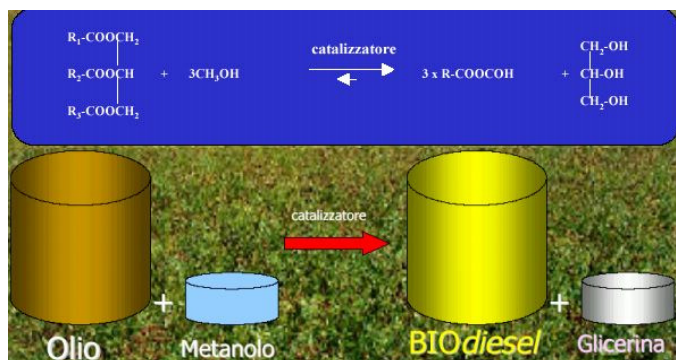


ESTRAZIONE DEI LIPIDI

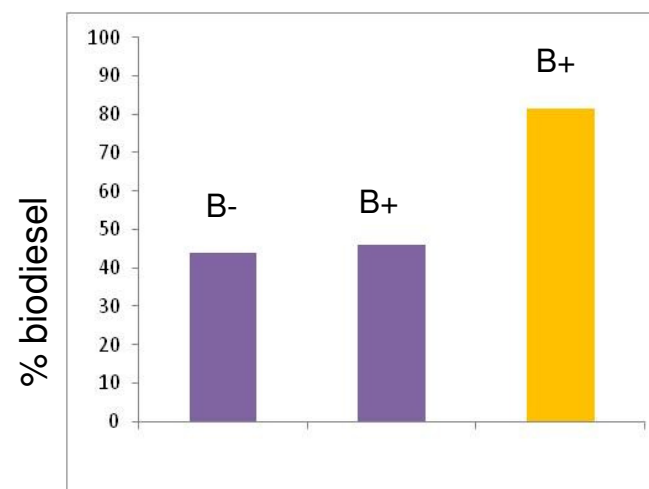
CLOROFORMIO : METANOLO
2 : 1



TRANSESTERIFICAZIONE DEGLI ACIDI GRASSI



N. oculata
Coccol green.



Marine and freshwater microalgae species	Lipid content (% dry weight biomass)	Lipid productivity (mg/L/day)	Volumetric productivity of biomass (g/L/day)	Areal productivity of biomass (g/m ² /day)
<i>Ankistrodesmus</i> sp.	24.0–31.0	–	–	11.5–17.4
<i>Botryococcus braunii</i>	25.0–75.0	–	0.02	3.0
<i>Chaetoceros muelleri</i>	33.6	21.8	0.07	–
<i>Chaetoceros calcitrans</i>	14.6–16.4/39.8	17.6	0.04	–
<i>Chlorella emersonii</i>	25.0–63.0	10.3–50.0	0.036–0.041	0.91–0.97
<i>Chlorella protothecoides</i>	14.6–57.8	1214	2.00–7.70	–
<i>Chlorella sorokiniana</i>	19.0–22.0	44.7	0.23–1.47	–
<i>Chlorella vulgaris</i>	5.0–58.0	11.2–40.0	0.02–0.20	0.57–0.95
<i>Chlorella</i> sp.	10.0–48.0	42.1	0.02–2.5	1.61–16.47/25
<i>Chlorella pyrenoidosa</i>	2.0	–	2.90–3.64	72.5/130
<i>Chlorella</i>	18.0–57.0	18.7	–	3.50–13.90
<i>Chlorococcum</i> sp.	19.3	53.7	0.28	–
<i>Cryptocodinium cohnii</i>	20.0–51.1	–	10	–
<i>Dunaliella salina</i>	6.0–25.0	116.0	0.22–0.34	1.6–3.5/20–38
<i>Dunaliella primolecta</i>	23.1	–	0.09	14
<i>Dunaliella tertiolecta</i>	16.7–71.0	–	0.12	–
<i>Dunaliella</i> sp.	17.5–67.0	33.5	–	–
<i>Ellipsoidion</i> sp.	27.4	47.3	0.17	–
<i>Euglena gracilis</i>	14.0–20.0	–	7.70	–
<i>Haematococcus pluvialis</i>	25.0	–	0.05–0.06	10.2–36.4
<i>Isochrysis galbana</i>	7.0–40.0	–	0.32–1.60	–
<i>Isochrysis</i> sp.	7.1–33	37.8	0.08–0.17	–
<i>Monodus subterraneus</i>	16.0	30.4	0.19	–
<i>Monallanthus salina</i>	20.0–22.0	–	0.08	12
<i>Nannochloris</i> sp.	20.0–56.0	60.9–76.5	0.17–0.51	–
<i>Nannochloropsis oculata</i>	22.7–29.7	84.0–142.0	0.37–0.48	–
<i>Nannochloropsis</i> sp.	12.0–53.0	37.6–90.0	0.17–1.43	1.9–5.3
<i>Neochloris oleoabundans</i>	29.0–65.0	90.0–134.0	–	–
<i>Nitzschia</i> sp.	16.0–47.0	–	–	8.8–21.6
<i>Oocystis pusilla</i>	10.5	–	–	40.6–45.8
<i>Pavlova salina</i>	30.9	49.4	0.16	–
<i>Pavlova lutheri</i>	35.5	40.2	0.14	–
<i>Phaeodactylum tricornutum</i>	18.0–57.0	44.8	0.003–1.9	2.4–21
<i>Porphyridium cruentum</i>	9.0–18.8/60.7	34.8	0.36–1.50	25
<i>Scenedesmus obliquus</i>	11.0–55.0	–	0.004–0.74	–
<i>Scenedesmus quadricauda</i>	1.9–18.4	35.1	0.19	–
<i>Scenedesmus</i> sp.	19.6–21.1	40.8–53.9	0.03–0.26	2.43–13.52
<i>Skeletonema</i> sp.	13.3–31.8	27.3	0.09	–
<i>Skeletonema costatum</i>	13.5–51.3	17.4	0.08	–
<i>Spirulina platensis</i>	4.0–16.6	–	0.06–4.3	1.5–14.5/24–51
<i>Spirulina maxima</i>	4.0–9.0	–	0.21–0.25	25
<i>Thalassiosira pseudonana</i>	20.6	17.4	0.08	–
<i>Tetraselmis suecica</i>	8.5–23.0	27.0–36.4	0.12–0.32	19
<i>Tetraselmis</i> sp.	12.6–14.7	43.4	0.30	–

Mata et al. 2010

Table 1
Comparison of some sources of biodiesel

Chisti 2007

Crop	Oil yield (L/ha)	Land area needed (M ha) ^a	Percent of existing US cropping area ^a
Com	172	1540	846
Soybean	446	594	326
Canola	1190	223	122
Jatropha	1892	140	77
Coconut	2689	99	54
Oil palm	5950	45	24
Microalgae ^b	136,900	2	1.1
Microalgae ^c	58,700	4.5	2.5

^a For meeting 50% of all transport fuel needs of the United States.

^b 70% oil (by wt) in biomass.

^c 30% oil (by wt) in biomass.